

Shape- Based Tracking and Analysis of Myocardial Function from 4D Images

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Abstract

This paper is a status report of work-in-progress, funded by the National Heart, Lung and Blood Institute (NHLBI) of the National Institutes of Health (NIH), that is aimed at more accurately and objectively determining and quantifying the regional and global function of the left ventricle (LV) of the heart under both normal and ischemic conditions, fundamentally relying on computer vision-like models and strategies.

1 Introduction

The quantification of left ventricular (LV) regional function from diagnostic images permits clinically important measurements to be made that are crucial for managing patients with ischemic heart disease. However, such measurements have been hampered by limitations in conventional imaging and image analysis methodology that have commonly been applied in the clinical arena to date. These limitations have prohibited the accurate regional assessment of myocardial injury and/or the prediction of vessel patency, and include the facts that: 1.) most conventional imaging methods provide two-dimensional (2D) temporal slice or projection sequences (e.g. see [4]), that cannot possibly permit viewing or analysis of the true motion of the heart, which rigidly moves and non-rigidly deforms in a three-dimensional (3D) space. 2.) Most approaches to regional image-based analysis of LV function: a.) make gross and restrictive assumptions about the general direction of LV motion or thickening (e.g. towards a center of mass) and b.) utilize only the end diastolic (ED) and end systolic (ES) image frames (e.g. [3]), ignoring the fact that the LV actually goes through a temporal wave of contraction and that the asynchrony of surface motion: LV thickening from region to region may be indicative of ischemia.

Our current research utilizes techniques from computer vision to analyze 3D cardiac image sequences in order to more accurately estimate regional LV function, and is much more adaptable to the nonlinear, non-rigid regional motion of the LV than the techniques based

on the restrictive assumptions mentioned above. The approach is based on the use of non-invasive, three-dimensional (3D), cardiac diagnostic imaging sequences (i.e. four-dimensional (4D) data) acquired from any one of a number of modalities- currently 3D cine Computed Tomography (CT) acquired from the Dynamic Spatial Reconstructor (DSR) at the Mayo Clinic and cardiac Magnetic Resonance Image (MRI) data acquired at our own facility and soon to be tested on 3D transesophageal echocardiographic data. Our image analysis methodology follows the shape properties of the endocardial and epicardial surfaces of the LV over the entire cardiac cycle, based on locating and matching differential geometric landmark features and using a mathematical optimization reasoning strategy to combine smoothness models with data-derived information. In some respects, this approach has similar goals as other recent efforts in the medical imaging community that utilize the unique characteristics of MR imaging techniques to track cardiac wall motion and function: MR tagging (e.g. [2]) and phase-contrast velocity imaging ([6]). It is of interest to note that the shape-based approach can derive quantitative parameters of LV function similar to those derived from these MRI-based approaches, yet is not restricted to using only MRI data.

2 Image Analysis Methods

Surface segmentation. The 4D image datasets are first segmented by treating the data as if it consisted of a sequence of temporal frames where there exists a spatial stack of 2D images in each frame. We solve the 2D boundary finding problem twice in each image, once for the epicardial border and once for the endocardial border using a deformable contour approach that we have developed for 2D boundary finding [7]. The boundaries found in this plane are now used as a bias and initial estimate for locating the endocardial and epicardial boundaries in the next plane in the stack. This process repeats until all of the contours that make up the LV surfaces in each frame are completely located. Finally, the contours that form the endocardial and epicardial surface in each 3D frame are stacked and then sent to a surface

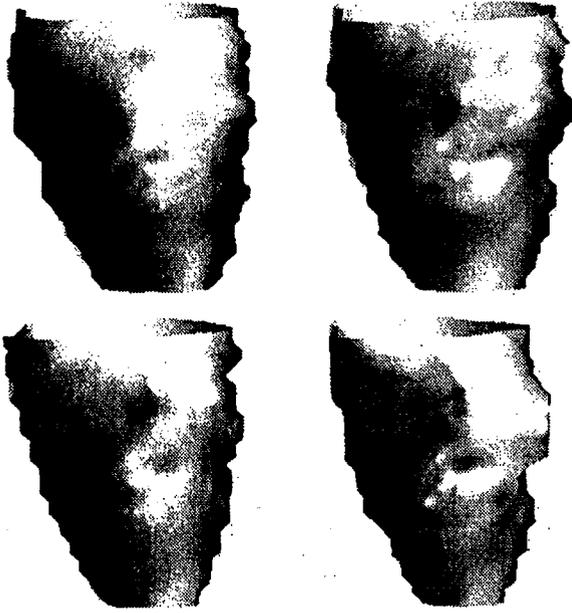


Figure 1: Gray scale renderings of color-coded bending energy maps of four, temporally-consecutive endocardial surfaces of the dog heart in its normal, or baseline, state. The sequence progresses as follows: upper left, upper right, lower left, lower right.

tessellation algorithm that we have implemented, which typically creates roughly 15,000 triangles per DSR-derived endocardial surface and 3,000 triangles per MRI-derived endocardial surface.

Shape-based motion tracking. As mentioned above, the movement over time of a set of LV surface points is performed by following local surface shape. Our approach reflects further developments, improvements and modifications to our earlier efforts in this area [1]. The dense set of points found from tessellating the surfaces are used to guide shape calculations. Between any 2 surfaces, shape matching is first performed through the use of a metric based on comparing surface patches using bending energy. The basic idea is to match a surface patch on a 3D (endocardial or epicardial) surface found from an image frame at time t with a surface patch within a region of surface patches on a corresponding 3D surface found from an image frame at time $t + \delta t$, using measurements of the patches' mean and Gaussian curvatures. We model the surface patches as elastic thin-plates, where the energy needed to bend each plate away from any arbitrarily formed initial surface to a deformed state is

$$\epsilon_{be}(u, v) = \left\{ \left[\frac{\kappa_1^2(u, v) + \kappa_2^2(u, v)}{2} \right]^{\frac{1}{2}} - \left[\frac{\bar{\kappa}_1^2(\bar{u}, \bar{v}) + \bar{\kappa}_2^2(\bar{u}, \bar{v})}{2} \right]^{\frac{1}{2}} \right\}^2 \quad (1)$$

The principal curvatures of a surface patch at time t_0 are given with no bars, while the same parameters at time $t_0 + \delta t$ (within a search window on the second surface) are specified with bars. This equation arrives at a numerical value measuring the energy that was required to deform the surface patch surrounding surface point (u, v) at time t_0 to achieve a particular shape at a test position within a search window at time $t_0 + \delta t$. Each of the $\frac{\kappa_1(u, v)^2 + \kappa_2(u, v)^2}{2}$ terms in this equation can separately be seen as computing the amount of bending energy required to bend a thin, flexible, flat plate to the shape of the surface patch surrounding point (u, v) . Pseudocolor maps of the local bending energies can be displayed on top of surface renderings of the LV. A gray scale display of such mappings/ renderings is shown in figure 1 for four sequential endocardial surfaces, reconstructed from a MRI 4D acquisition. The result of the matching process specified by equation (1) essentially compares the maps displayed in figure 1 and yields a set of initial shape-based, best-match vectors, $D_0(u, v)$, for pairs of surfaces derived from the 4D image sequence, as well as information about the uniqueness of the match within each search region. This information is now sent to a regularizing functional that will include terms to smooth over (u, v) surface space:

$$D^*(u, v) = \arg \min_{\mathbf{D}} \int_U \int_V C_{\mathbf{D}}(u, v) [D(u, v) - D_0(u, v) + \frac{\partial D(u, v)}{\partial \mathbf{u}}] d\mathbf{u}$$

In this equation, U, V is the domain of the surface at time t in which the finite element grid from the Delaunay triangulation is embedded, $\mathbf{u} = [uv]^T$ is the position of the end of the vector $D(u, v)$ on the surface found at time t , $D^*(u, v)$ is the optimal smoothed motion vector field between any two surfaces found at times t and $t + \delta t$, $D_0(u, v)$ is the initial motion vector estimate, and $C_{\mathbf{D}}(u, v)$ is the confidence measure matrix from the initial estimation, which weights the overall goodness of the initial match as well as the match uniqueness within the search area. While this model currently serves as a spatial smoothness model for the vector field, the bulk of our current efforts are aimed at incorporating appropriate temporal models as well. The solution of this functional will yield a set of smoothed flow vectors that better estimate true local LV motion. It should be clear that ultimately strong, unique matches, according to the bending energy match measure, that last over several frames, are weighted via the confidence measures to be the most influential shape-based landmarks. Flow vectors running through the knot points of the surface patches are connected in order to form trajectories that track individual points' motion over the entire cardiac cycle. The current strategy is to estimate and track the trajectories of motion of a dense field of points that sample the LV endocardial and epicardial surfaces at 1 point in time, over the entire cardiac cycle. The results

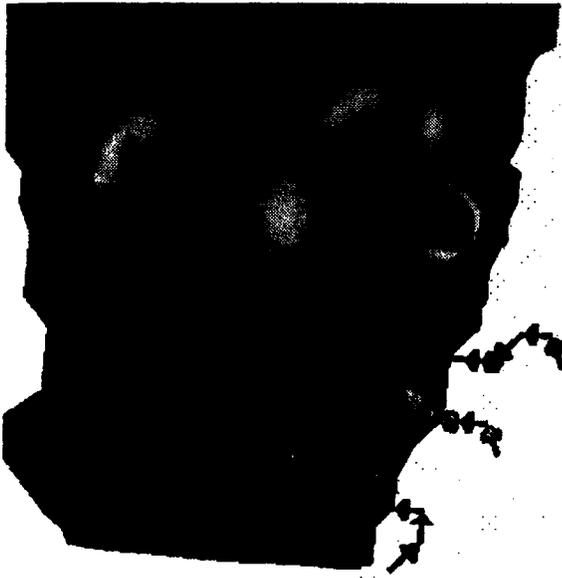


Figure 2: Algorithm- computed trajectories of several endocardial points for the normal or baseline heart. The trajectories begin at the end- diastolic surface and end at the end- systolic surface (rendered here).

Computing several of these trajectories in the anteropical region of the heart from MRI- derived endocardial surfaces, are shown in figure 2. LV motion and function indices are then derived from these trajectories, and include i.) the sum of the magnitudes (lengths) of the flow vectors, or path length $L_{path,\alpha}(h, i)$, ii.) the shape of the path (compared to a linear trajectory), $s_{path,\alpha}(h, i)$ and iii.) the peak velocity within the path, $v_{peak,\alpha}(h, i)$. Each of these measures are recorded for each trajectory, indexed by i , on each heart being studied, indexed by h . In general, these trajectories will be used to create quantitative measures of endocardial surface motion and LV thickening/ strain.

3 Experiments and Results

The methodology is currently being validated using computer simulations of linearly deformable objects and image data acquired from acute dog studies. The acute dogs will have image-distinguishable markers (radioopaque wires for cine CT and a combination of copper plugs (endocardium) and a gadolinium/ water mixture- lled pellet (epicardium) for MRI) sewn to the LV wall.

Images from 3 slices of a single 3D frame from a 4D MRI acquisition are shown in figure 3. The arrows point to the markers (dark voids on the endocardium and bright spots on the epicardium). The validation testing is being performed utilizing data from 5 studies acquired from each of two different imaging modalities (i.e. the current test set consists of ten 4D data sets): 4D gated/cine Magnetic Resonance Imaging and 4D real-

time cine Computed Tomographic Imaging (using data from the Dynamic Spatial Reconstructor). Table 1 illustrates the comparison of bending energy-derived measures from an infarcted heart derived from DSR image data to the same measures derived from the manually tracked movement of four implanted endocardial markers (i.e. the gold standard) derived from the same images. These comparisons were made using only the raw, unsmoothed curviness vectors $D_0(u, v)$ to perform the matching from which the algorithm- based trajectories were derived.

Ultimately, our goal is to better distinguish and characterize the spatial/ temporal extent and function of regions of ischemia and infarction in the LV wall, as well as gain insight into LV post- MI remodeling. Thus, further studies are underway to compare these algorithm- derived *in vivo* measures to measures of *post mortem* injury. By studying a spatially dense set of points that sample the LV wall's surfaces over many temporal frames within a cardiac cycle and over a range of conditions of LV wall abnormality, we will be able to better understand the regional and global physiological processes associated with ischemic heart disease.

4 Summary

In the workshop presentation of this paper, we have described a shape- based LV motion tracking algorithm, the image data we are using, and the graphical methods being used to visualize the results. In addition, we have presented initial qualitative and quantitative data aimed at comparing the algorithm- derived motion trajectories to those found using implanted marker gold standards. Further methodology work continues along the lines of developing improved temporal models for motion tracking, including incorporating additional differential geometric features into the matching algorithm and forming more complete 4D visualization approaches. Validation and clinical experimentation continues based on the *in vivo* acute infarct models and using DSR and MR image data.

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Figure 3: Three slices from the same 3D MRI time frame, illustrating the paired markers used for motion tracking validation (white arrows= epicardial markers, black= endocardial markers).

Marker no.	path length			displacement length		
	$L_{path,gold}$	$L_{path,alg}$	δ_{path}	$L_{disp,gold}$	$L_{disp,alg}$	δ_{disp}
1	30.48	29.77	0.71	12.02	15.44	3.42
2	36.38	25.28	11.10	17.47	15.27	2.20
3	23.89	31.65	7.76	13.64	12.02	1.62
4	32.15	26.48	5.67	12.42	12.44	0.02
μ_{δ}			6.31mm			1.81mm
σ_{δ}			3.77mm			1.22mm

Table 1: Table of path lengths ($L_{path,\alpha}$) and complete path displacements ($L_{disp,\alpha}$) at four endocardial marker positions of a single heart. These data were computed using 1mm³ 4D DSR image data, and compare algorithm-computed trajectories ($\alpha = alg$) to the trajectories of implanted markers ($\alpha = gold$). Note that μ_{δ} and σ_{δ} represent the means and standard deviations of the differences in the measurements.

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